Package 'pram'

November 15, 2024

Title Pooling RNA-seq datasets for assembling transcript models

Version 1.22.0

Description Publicly available RNA-seq data is routinely used for retrospective analysis to elucidate new biology. Novel transcript discovery enabled by large collections of RNA-seq datasets has emerged as one of such analysis. To increase the power of transcript discovery from large collections of RNA-seq datasets, we developed a new R package named Pooling RNA-seq and Assembling Models (PRAM), which builds transcript models in intergenic regions from pooled RNA-seq datasets. This package includes functions for defining intergenic regions, extracting and pooling related RNA-seq alignments, predicting, selected, and evaluating transcript models.

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Encoding UTF-8

LazyData true

URL https://github.com/pliu55/pram

BugReports https://github.com/pliu55/pram/issues

Depends R (>= 3.6)

Imports methods, BiocParallel, tools, utils, data.table (>= 1.11.8), GenomicAlignments (>= 1.16.0), rtracklayer (>= 1.40.6), BiocGenerics (>= 0.26.0), GenomeInfoDb (>= 1.16.0), GenomicRanges (>= 1.32.0), IRanges (>= 2.14.12), Rsamtools (>= 1.32.3), S4Vectors (>= 0.18.3)

RoxygenNote 6.1.0

Suggests testthat, BiocStyle, knitr, rmarkdown

Collate 'Param.R' 'Transcript.R' 'buildModel.R' 'defIgRanges.R' 'evalModel.R' 'prepIgBam.R' 'runPRAM.R' 'selModel.R' 'util.R'

VignetteBuilder knitr

biocViews Software, Technology, Sequencing, RNASeq, BiologicalQuestion, GenePrediction, GenomeAnnotation, ResearchField, Transcriptomics

SystemsRequirements buildModel() and runPRAM() functions require external software Cufflinks, StringTie, and/or TACO. For details, please see the 'Required external software' section in vignette's 'Building transcript models: buildModel()'.

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Author Peng Liu [aut, cre],
Colin N. Dewey [aut],
Sündüz Keleş [aut]

Maintainer Peng Liu <pliu55.wisc+bioconductor@gmail.com>

Contents

`buildModel` *Build transcript models from aligned RNA-seq data*

Description

Build transcript models from aligned RNA-seq data

Usage

```
buildModel(in_bamv, out_gtf, method = "plcf", nthreads = 1,
    tmpdir = NULL, keep_tmpdir = FALSE, cufflinks = "",
    stringtie = "", taco = "", cufflinks_ref_fa = "")
```

Arguments

`in_bamv`	A character vector of input BAM file(s). If mode 'cf' or 'st' is used, only one input RNA-seq BAM file is allowed. Currently, PRAM only supports strand-specific paired-end data with the first mate on the right-most of transcript coordinate, i.e., 'fr-firststrand' by Cufflinks's definition.
`out_gtf`	An output GTF file of predicted transcript models
`method`	A character string defining PRAM's model building method. Current available methods are: • plcf: pooling + cufflinks • plst: pooling + stringtie • cfmg: cufflinks + cuffmerge • stmg: stringtie + merging

- cftc: cufflinks + taco
- cf: cufflinks
- st: stringtie

Default: 'plcf'

`nthreads` An integer defining the number of threads to-be-used. Default: 1

`tmpdir` A character string defining the full name of a folder for saving temporary files. If not tmpdir is give, PRAM will use R's tempdir().

`keep_tmpdir` Whether to keep temporary files afterwards. Default: False

`cufflinks` Cufflinks executable. Required by mode 'plcf', 'cfmg', and 'cf'. For mode 'cfmg', executable files of Cuffmerge, Cuffcompare, and gtf_to_sam from the Cufflinks suite are assumed to be under the same folder as Cufflinks. All the executables are available to download for Linux http://cole-trapnell-lab.github.io/cufflinks/assets/downloads/cufflinks-2.2.1.Linux_x86_64.tar.gz and MacOS http://cole-trapnell-lab.github.io/cufflinks/assets/downloads/cufflinks-2.1.1.OSX_x86_64.tar.gz. Souce code can be obtained from http://cole-trapnell-lab.github.io/cufflinks/. Default: ''

`stringtie` StringTie executable file. Required by mode 'plst', 'stmg', and 'st'. Executable can be downloaded for Linux http://ccb.jhu.edu/software/stringtie/dl/stringtie-1.3.3b.Linux_x86_64.tar.gz and MacOS http://ccb.jhu.edu/software/stringtie/dl/stringtie-1.3.3b.OSX_x86_64.tar.gz. Souce code can be obtained from https://ccb.jhu.edu/software/stringtie/. Default: ''

`taco` TACO executable file. Required by mode 'cftc'. Executable can be downloaded for Linux https://github.com/tacorna/taco/releases/download/v0.7.0/taco-v0.7.0.Linux_x86_64.tar.gz and MacOS https://github.com/tacorna/taco/releases/download/v0.7.0/taco-v0.7.0.OSX_x86_64.tar.gz. Souce code can be obtained from https://tacorna.github.io. Default: ''

`cufflinks_ref_fa` Genome reference fasta file for Cufflinks. If supplied, will be used for cufflinks's '–frag-bias-correct' and cuffmerge's '–ref-sequence' options. Default: ''

Value

None

Examples

```
fbams = c(  system.file('extdata/bam/CMPRep1.sortedByCoord.clean.bam',
                        package='pram'),
            system.file('extdata/bam/CMPRep2.sortedByCoord.clean.bam',
                        package='pram') )

foutgtf = tempfile(fileext='.gtf')

## assuming the stringtie binary is in folder /usr/local/stringtie-1.3.3/
## you can run buildModel() by the following example
##
# buildModel(fbams, foutgtf, method='plst',
#            stringtie='/usr/local/stringtie-1.3.3/stringtie')
```

defIgRanges *Define intergenic genomic regions*

Description

Define intergenic genomic regions

Usage

```
defIgRanges(in_gtf, chromgrs, genome = NULL, fchromsize = NULL,
    radius = 10000, feat = "exon", chroms = NULL)
```

Arguments

in_gtf	An input GTF file for defining genomic coordinates of existing genes. Required to have 'gene_id' in the attribute column (column 9)
chromgrs	A GRanges object defining chromosome sizes.
genome	Version of the genome. Will be used when 'chromgrs' is missing. Currently supported ones are: • hg19 • hg38 • mm9 • mm10 All the above genomes have sizes for all chromosomes including random and alt ones. Default: NULL
fchromsize	Name of a file defining chromosome sizes. Will be used when 'chromgrs' and 'genome' are missing. It can be downloaded from UCSC, e.g. for hg19, http://hgdownload.cse.ucsc.edu/ goldenpath/hg19/database/chromInfo.txt.gz Required to have at least two tab-delimited columns without any header: 1. chromosome name, e.g. chr1 2. chromosome length, e.g. 249250621 Both uncompressed and gzipped files are supported. Default: NULL
radius	Region length (bp) of gene's upstream and downstream to be excluded from intergenic region. Default: 10,000
feat	Feature in the GTF file (column 3) to-be-used for defining genic region. Default: exon
chroms	A vector of chromosomes names to define intergenic regions. e.g. c('chr10', 'chr11') Default: NULL

Value

a GRanges object of intergenic regions

Examples

```
fgtf = system.file('extdata/gtf/defIgRanges_in.gtf', package='pram')


defIgRanges(fgtf, genome='hg38')
```

`evalModel` *Evaluate transcript model*

Description

Evaluate trascript model's precision and recall on exon nucleotides, splice junctions, and splice patterns by comparing them to transcript targets

Usage

```
evalModel(model_exons, target_exons)

## S4 method for signature 'GRanges,GRanges'
evalModel(model_exons, target_exons)

## S4 method for signature 'character,character'
evalModel(model_exons, target_exons)

## S4 method for signature 'data.table,data.table'
evalModel(model_exons, target_exons)

## S4 method for signature 'character,data.table'
evalModel(model_exons, target_exons)
```

Arguments

`model_exons`	genomic coordinates for transcript model exons
`target_exons`	genomic coordinates for transcript target exons

Value

a data table of precision, recall, number of true positive, false negative, false positive for all three evaluated features

Methods (by class)

- `model_exons = GRanges, target_exons = GRanges`: Both **model_exons** and **target_exons** are GRanges objects to define genomic coordinates of exons. Required to have a meta-data column named 'trid' to define each exon's transcript ID.
- `model_exons = character, target_exons = character`: Both **model_exons** and **target_exons** are GTF files with full names. Each GTF file is required to have a 'transcript_id' tag in column 9.

- `model_exons = data.table, target_exons = data.table`: Both **model_exons** and **target_exons** are data.table objects to define exon genomic coordinatess. Required to have the following columns:
  - chrom: exon's chromosome, e.g. 'chr8'
  - start: exon's start position
  - end: exon's end position
  - strand: exon's strand, '+' or '-'
  - trid: exon's transcript ID
- `model_exons = character, target_exons = data.table`: The **model_exons** is a GTF file with full name and **target_exons** is a data.table object. Requirements for GTF and data.table are the same as above

Examples

```
fmdl = system.file('extdata/benchmark/plcf.tsv', package='pram')
ftgt = system.file('extdata/benchmark/tgt.tsv',  package='pram')

mdldt = data.table::fread(fmdl, header=TRUE, sep="\t")
tgtdt = data.table::fread(ftgt, header=TRUE, sep="\t")


evalModel(mdldt, tgtdt)
```

---

prepIgBam *Extract alignments in intergenic regions from BAM files*

Description

Extract alignments in intergenic regions from BAM files

Usage

```
prepIgBam(finbam, iggrs, foutbam, max_uni_n_dup_aln = 10,
    max_mul_n_dup_aln = 10)
```

Arguments

`finbam`	Full name of an input RNA-seq BAM file. Currently, PRAM only supports strand-specific paired-end data with the first mate on the right-most of transcript coordinate, i.e., 'fr-firststrand' by Cufflinks's definition
`iggrs`	A GenomicRanges object defining intergenic regions
`foutbam`	Full name of an output BAM file to save all alignment fell into intergenic regions
`max_uni_n_dup_aln`	Maximum number of uniquely mapped fragments to report per each alignment. Default: 10
`max_mul_n_dup_aln`	Maximum number of multi-mapping fragments to report per each alignment. Default: 10

Value

None

Examples

```
finbam = system.file('extdata/bam/CMPRep2.sortedByCoord.raw.bam',
                     package='pram')

iggrs = GenomicRanges::GRanges('chr10:77236000-77247000:+')

foutbam = tempfile(fileext='.bam')


prepIgBam(finbam, iggrs, foutbam)
```

---

runPRAM *Predict intergenic transcript models from RNA-seq*

Description

Predict intergenic transcript models from RNA-seq

Usage

```
runPRAM(in_gtf, in_bamv, out_gtf, method, cufflinks = "",
    stringtie = "", taco = "")
```

Arguments

in_gtf	An input GTF file for defining genomic coordinates of existing genes. Required to have 'gene_id' in the attribute column (column 9)
in_bamv	A character vector of input BAM file(s). If mode 'cf' or 'st' is used, only one input RNA-seq BAM file is allowed. Currently, PRAM only supports strand-specific paired-end data with the first mate on the right-most of transcript coordinate, i.e., 'fr-firststrand' by Cufflinks's definition.
out_gtf	An output GTF file of predicted transcript models
method	A character string defining PRAM's model building method. Current available methods are: • plcf: pooling + cufflinks • plst: pooling + stringtie • cfmg: cufflinks + cuffmerge • stmg: stringtie + merging • cftc: cufflinks + taco • cf: cufflinks • st: stringtie Default: 'plcf'

cufflinks — Cufflinks executable. Required by mode 'plcf', 'cfmg', and 'cf'. For mode 'cfmg', executable files of Cuffmerge, Cuffcompare, and gtf_to_sam from the Cufflinks suite are assumed to be under the same folder as Cufflinks. All the executables are available to download for Linux http://cole-trapnell-lab.github.io/cufflinks/assets/downloads/cufflinks-2.2.1.Linux_x86_64.tar.gz and MacOS http://cole-trapnell-lab.github.io/cufflinks/assets/downloads/cufflinks-2.1.1.OSX_x86_64.tar.gz. Souce code can be obtained from http://cole-trapnell-lab.github.io/cufflinks/. Default: ''

stringtie — StringTie executable file. Required by mode 'plst', 'stmg', and 'st'. Executable can be downloaded for Linux http://ccb.jhu.edu/software/stringtie/dl/stringtie-1.3.3b.Linux_x86_64.tar.gz and MacOS http://ccb.jhu.edu/software/stringtie/dl/stringtie-1.3.3b.OSX_x86_64.tar.gz. Souce code can be obtained from https://ccb.jhu.edu/software/stringtie/. Default: ''

taco — TACO executable file. Required by mode 'cftc'. Executable can be downloaded for Linux https://github.com/tacorna/taco/releases/download/v0.7.0/taco-v0.7.0.Linux_x86_64.tar.gz and MacOS https://github.com/tacorna/taco/releases/download/v0.7.0/taco-v0.7.0.OSX_x86_64.tar.gz. Souce code can be obtained from https://tacorna.github.io. Default: ''

Value

None

Examples

```
in_gtf = system.file('extdata/demo/in.gtf', package='pram')

in_bamv = c(system.file('extdata/demo/SZP.bam', package='pram'),
            system.file('extdata/demo/TLC.bam', package='pram') )

pred_out_gtf = tempfile(fileext='.gtf')

## assuming the stringtie binary is in folder /usr/local/stringtie-1.3.3/
## you can run runPRAM() by the following example
##
# runPRAM(in_gtf, in_bamv, pred_out_gtf, method='plst',
#         stringtie='/usr/local/stringtie-1.3.3/stringtie')
```

---

selModel *Select transcript models*

Description

Select transcript models

Usage

```
selModel(fin_gtf, fout_gtf, min_n_exon = 2, min_tr_len = 200,
    info_keys = c("transcript_id"))
```

Arguments

fin_gtf	Character of an input GTF file that contains transcript models. Required to have 'transcript_id' in the attribute column (column 9)
fout_gtf	Character of an output GTF file that contains selected transcript models
min_n_exon	Minimium number of exons a transcript model required to have Default: 2
min_tr_len	Minimium length (bp) of exon(s) and intron(s) a transcript model required to have Default: 200
info_keys	A vector of characters defining the attributes in input GTF file's column 9 to be saved in the output GTF file. 'transcript_id' will always be saved. Default: c('transcript_id')

Value

None

Examples

```
fin_gtf = system.file('extdata/gtf/selModel_in.gtf', package='pram')

fout_gtf = tempfile(fileext='.gtf')


selModel(fin_gtf, fout_gtf)
```

Index